Store at -20°C

SimpleChIP[®] Human CD11b **Promoter Primers**

coated particles.

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Entrez-Gene ID #3684

UniProt ID #P11215

New 07/14

1.5

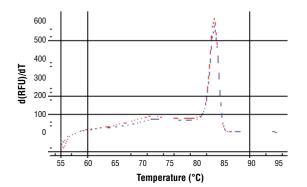
For Research Use Only. Not For Use In Diagnostic Procedures.

	Applications	Species Cross-Reactivity*	Primer Anneal/Exte	nsion PCR Product Length
	ChIP	H	65°C	107 bp
contair specifi molecu to amp munop use in tested IP Kits	n a mix of forward and reve c to a region of the human ule 11b (CD11b) promoter. olify DNA that has been isol precipitation (ChIP). Primer SYBR® Green quantitative in conjunction with Simple #9002 and #9003 and ChI	cluster of differentiation These primers can be used ated using chromatin im-	(°) 32 30 28 24 22 -2 -1.5 Lo	1 -0.5 0 0.5 1.0 g Starting Quantity (ng)

SimpleChIP® Human CD11b Promoter Primers were tested on DNA isolated from cross-linked cells using the SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. Real-time PCR was performed in duplicate on a serial dilution of 2% total input DNA (20 ng, 4 ng, 0.8 ng, and 0.16 ng) using a real-time PCR detection system and SYBR® Green reaction mix. The PCR amplification efficiency (E) and correlation coefficient (R²) were calculated based on the corresponding threshold cycle (C_{τ}) of each dilution sample during 40 cycles of real-time PCR (95°C denaturation for 15 sec, 65°C anneal/ extension for 60 sec).

F= 102 7%

B²= 0 991



antigen-like family member B (integrin alpha-M), a cell surface

receptor involved in adhesion to various lymphocytes and in

binding to complement C3b, mediating uptake of complement-

PCR product melting curves were obtained for real-time PCR reactions performed using SimpleChIP® Human CD11b Promoter Primers. Data is shown for both duplicate PCR reactions using 20 ng of total DNA. The melt curve consists of 80 melt cycles, starting at 55°C with increments of 0.5°C per cycle. Each peak is formed from the degradation of a single PCR product.

Directions for Use: 1. Label the appropriate number of PCR tubes or PCR plates

at -20°C.

compatible with the model of real-time PCR machine to be used. PCR reactions should be performed in duplicate and should include a tube with no DNA to control for contamination, and a serial dilution of a 2% total input chromatin DNA (undiluted, 1:5, 1:25, 1:125), which is used to create a standard curve and determine amplification efficiency.

Storage: Supplied in nuclease-free water at a concentration of 5 μ M (each primer is at a final concentration of 5 μ M). Store

2. Add 2 µl of the appropriate ChIP DNA sample to each tube or well of the PCR plate.

3. Prepare a master PCR reaction mix as described below. Add enough reagents for two extra reactions to account for loss of volume. Add 18 μ l of the master PCR reaction mix to each PCR reaction tube or well of the PCR plate.

Reagent	Volume for 1 PCR Reac	tion (20 µl)
Nuclease-free H ₂ O		6 µl
5 µM SimpleChIP® Primers		2 µl
2X SYBR® Green Re	action Mix	10 µl

4. Start the following PCR reaction program:

a. Initial Denaturation: 95°C for 3 min.

- b. Denaturation: 95°C for 15 sec.
- c. Anneal and Extension: Primer-specific temp. for 60 sec.
- d. Repeat steps b and c for a total of 40 cycles.

5. Analyze quantitative PCR results using software provided with the real-time PCR machine.

For product specific protocols and a complete listing of recommended companion products please see the product web page at www.cellsignal.com

SYBR® Green is a registered trademark of Life Technologies Corporation.

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Applications: W—Western IP—Immunoprecipitation IHC—Immunohistochemistry ChIP—Chromatin Immunoprecipitation IF—Immunofluorescence F—Flow cytometry E-P—ELISA-Peptide Species Cross-Reactivity: H—human M—mouse R—rat Hm—hamster Mk—monkey Mi—mink C—chicken Dm—D. melanogaster X—Xenopus Z—zebrafish B—bovine Dg—dog Pg—pig Sc—S. cerevisiae Ce—C. elegans Hr—Horse AII—all species expected Species enclosed in parentheses are predicted to react based on 100% homology