#31195

SimpleChIP[®] Mouse Pdk4 **Promoter Primers**

500 µl (250 PCR reactions) Cell Signaling

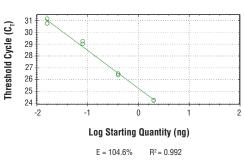
Support: +1-978-867-2388 (U.S.) www.cellsignal.com/support

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Entrez-Gene ID #5166 UniProt ID #Q16654

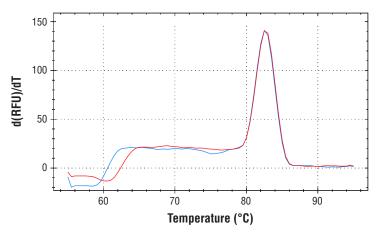
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Applications Species Cross-Reactivity **Primer Anneal/Extension** PCR Product Length ChIP 65°C М 127 bp Description: SimpleChIP® Mouse Pdk4 Promoter Primers 31 Threshold Cycle (C_T) contain a mix of forward and reverse PCR primers that are 30 specific to a region of the mouse pyruvate dehydrogenase ki-29 28 nase 4 (Pdk4) promoter. These primers can be used to amplify 27 DNA that has been isolated using chromatin immunoprecipi-26 tation (ChIP). Primers have been optimized for use in quantita-25 tive real-time PCR using SimpleChIP® Universal qPCR Master 24 Mix #88989. Primers have been tested in conjunction with SimpleChIP® Plus Enzymatic Chromatin IP Kits #9004 and #9005 and ChIP-validated antibodies from Cell Signaling Technology[®]. E = 104.6%



New 02/20

SimpleChIP® Mouse Pdk4 Promoter Primers were tested on DNA isolated from cross-linked cells using the SimpleChIP® Plus Enzymatic Chromatin IP Kit (Magnetic Beads) #9005. Real-time PCR was performed in duplicate on a serial dilution of 2% total input DNA (20 ng, 4 ng, 0.8 ng, and 0.16 ng) using a real-time PCR detection system and SimpleChIP® Universal qPCR Master Mix #88989. The PCR amplification efficiency (E) and correlation coefficient (R²) were calculated based on the corresponding threshold cycle (C_{τ}) of each dilution sample during 40 cycles of real-time PCR (95°C denaturation for 15 sec, 65°C anneal/extension for 60 sec).



PCR product melting curves were obtained for real-time PCR reactions performed using SimpleChIP® Mouse Pdk4 Promoter Primers. Data is shown for both duplicate PCR reactions using 20 ng of total DNA. The melt curve consists of 80 melt cycles, starting at 55°C with increments of 0.5°C per cycle. Each peak is formed from the degradation of a single PCR product.

Storage: Supplied in nuclease-free water at a concentration of 5 μ M (each primer is at a final concentration of 5 μ M). Store at -20°C.

Directions for Use:

- 1. Label the appropriate number of PCR tubes or PCR plates compatible with the model of real-time PCR machine to be used. PCR reactions should be performed in duplicate and should include a tube with no DNA to control for contamination, and a serial dilution of a 2% total input chromatin DNA (undiluted, 1:5, 1:25, 1:125), which is used to create a standard curve and determine amplification efficiency.
- 2. Add 2 µl of the appropriate ChIP DNA sample to each tube or well of the PCR plate.
- 3. Prepare a master PCR reaction mix as described below. Add enough reagents for two extra reactions to account for loss of volume. Add 18 µl of the master PCR reaction mix to each PCR reaction tube or well of the PCR plate.

Reagent Volume for 1 PCR Reaction	(20 µl)
Nuclease-free H ₂ O	6 µl
5 µM SimpleChĺP® Primers	2 µl
SimpleChIP® Universal qPCR Master Mix #88989	10 µl

4. Start the following PCR reaction program: a. Initial Denaturation: 95°C for 3 min.

- b. Denaturation: 95°C for 15 sec.
- c. Anneal and Extension: Primer-specific temp. for 60 sec.
- d. Repeat steps b and c for a total of 40 cycles.
- 5. Analyze guantitative PCR results using software provided with the real-time PCR machine.

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Applications: W—Western IP—Immunoprecipitation IHC—Immunohistochemistry ChIP—Chromatin Immunoprecipitation IF—Immunofluorescence F—Flow cytometry E-P—Fl ISA-Pentide Species Cross-Reactivity: H—human M—mouse R—rat Hm—hamster -monkey Mi-mink C-chicken Dm-D. melanogaster X-Xenopus Z-zebrafish B-bovine Dg-dog Pg-pig Sc-S. cerevisiae Ce-C. elegans Hr-Horse All-all species expected Species enclosed in parentheses are predicted to react based on 100% homology