Store at

10X Wash Buffer (CUT&RUN, CUT&Tag)



15 mL



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For Research Use Only. Not for Use in Diagnostic Procedures.

Description: The 10X Wash Buffer (CUT&RUN, CUT&Tag) provides enough reagent to support 24 CUT&RUN or CUT&Tag assays. This product is formulated for optimal performance in the CUT&RUN and CUT&Tag assays and each lot is tested and validated using the CUT&RUN Assay Kit #86652 or the CUT&Tag Assay Kit #77552. This product should be diluted to 1X using nuclease-free water and an appropriate amount of 100X Spermidine #27287 and Protease Inhibitor Cocktail (200X) #7012 should be added right before use. Please keep at room temperature during use to minimize stress on the cells.

Background: Like the chromatin immunoprecipitation (ChIP) assay, Cleavage Under Targets and Release Using Nuclease (CUT&RUN) and Cleavage Under Targets and Tagmentation (CUT&Tag) are powerful and versatile techniques used for probing protein-DNA interactions within the natural chromatin context of the cell (1-7). CUT&RUN provides a rapid, robust, and true low cell number assay for detection of protein-DNA interactions in the cell. Unlike the ChIP assay, CUT&RUN is free from formaldehyde cross-linking, chromatin fragmentation, and immunoprecipitation, making it a much faster and more efficient method for enriching protein-DNA interactions and identifying target genes. CUT&RUN can be performed in less than one day, from live cells to purified DNA, and has been shown to work with as few as 500-1,000 cells per assay (1,2). Instead of fragmenting all of the cellular chromatin as done in ChIP, CUT&RUN utilizes an antibodytargeted digestion of chromatin, resulting in much lower background signal than seen in the ChIP assay. As a result, CUT&RUN requires only 1/10th the sequencing depth that is required for ChIP-seq assays (1,2). Finally, the inclusion of simple spike-in control DNA allows for accurate quantification and normalization of target-protein binding that is not possible with the ChIP method. This provides for effective normalization of signals between samples and between experiments. CUT&Tag has many of the same advantages as the CUT&RUN assay in that it provides a rapid, robust, and true low cell number protocol for detection of protein-DNA interactions in the cell. In addition, the CUT&Tag assay adds an in situ adaptor DNA ligation step carried out by the pAG-Tn5 enzyme, in which an adaptor DNA is ligated directly to antibody-targeted chromatin DNA fragments in the cell. As a result, subsequent DNA library preparation is much faster and easier than library preparation following the CUT&RUN assay, free from DNA end repair, A-tailing, and adaptor ligation in vitro. CUT&Tag works very well for analyzing histone modifications, in addition to mapping some transcription factors and cofactors binding.

Storage: Store 10X Wash Buffer (CUT&RUN, CUT&Tag) at 4°C. This product is stable for at least 12 months.

Please visit www.cellsignal.com for a complete listing of recommended companion products.

Directions for Use: For the CUT&RUN and CUT&Tag assays, we recommend preparing 2 ml 1X Complete Wash Buffer for each cell line and an additional 100 μ l for each reaction or input sample. For example, to prepare 2.5 ml of 1X Complete Wash Buffer, add 250 μ l 10X Wash Buffer (CUT&RUN, CUT&Tag), 25 μ l 100X Spermidine #27287, and 12.5 μ l Protease Inhibitor Cocktail (200X) #7012 to 2,212.5 μ l nuclease-free water right before use. Equilibrate it to room temperature to minimize stress on the cells.

Background References:

- (1) Skene, P.J. and Henikoff, S. (2017) Elife 6, .
- (2) Skene, P.J. et al. (2018) Nat Protoc 13, 1006-1019.
- (3) Meers, M.P. et al. (2019) *Elife* 8, pii: e46314. doi: 10.7554/eLife.46314.
- (4) Meers, M.P. et al. (2019) Mol Cell 75, 562-575.e5.
- (5) Kaya-Okur, H.S. et al. (2019) Nat Commun 10, 1930.
- (6) Kaya-Okur, H.S. et al. (2020) Nat Protoc 15, 3264-3283.
- (7) Henikoff, S. et al. (2021) Bio Protoc 11, e4043.

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