Revision 2

e at +4C	PathScan [®] Total Akt1 Sandwich ELISA Antibody Pair				
Store		Orders: 877-616-CELL (2355) orders@cellsignal.com			
	1 Kit (Reagents for 4 x 96 well plates)	Support: 877-678-TECH (8324)			
7142	Species Cross Reactivity: UniProt ID: Entrez-Gene Id: H M #P31749 #207	Web: info@cellsignal.com cellsignal.com			
1		3 Trask Lane Danvers Massachusetts 01923 USA			
For Research Use Only. Not for Use in Diagnostic Procedures.					

Product Includes	Product #	Volume	Cap Color	Storage Temp
Akt Capture Rabbit mAb (100X)	90629	400 µl	Pink	+4C
Akt1 Detection Mouse mAb (100X)	20910	400 µl	Blue	+4C
Anti-mouse IgG, HRP-linked Antibody (1000X)	16736	40 µl	Yellow	-20C

Please visit cellsignal.com for a complete listing of recommended companion products.

Description	CST's PathScan [®] Total Akt1 Sandwich ELISA Antibody Pair is being offered as an economical alternative to our PathScan [®] Total Akt1 Sandwich ELISA Kit #7170. Capture and Detection antibodies (100X stocks) and HRP-conjugated secondary antibody (1000X stock) are supplied. Sufficient reagents are supplied for 4 x 96 well ELISAs. The Akt Rabbit Capture Antibody is coated in PBS overnight in a 96 well microplate. After blocking, cell lysates are added followed by Akt1 Mouse Detection Antibody and HRP- conjugated Anti-Mouse IgG. HRP substrate, TMB, is added for color development. The magnitude of the absorbance for this developed color is proportional to the quantity of Total Akt1 protein. Antibodies in kit are custom formulations specific to kit.
Reagents Not Supplied	Phosphate Buffered Saline (PBS-20X) #9808 Phosphate Buffered Saline with Tween-20 (PBST-20X) #9809 Cell Lysis Buffer (10X) #9803 TMB Substrate #7004 STOP Solution #7002 Blocking Buffer: 1X PBS/0.5% Tween-20, 1% BSA 96 Well Microplates** Microplate Reader ** Antibody Pairs have been validated on Corning© 96 Well Clear Polystyrene High Bind Stripwell [™] Microplates (#2592).
	Notes: Antibody pairs have been optimized using recommended buffers, reagents, plates and the included protocol. Solutions should be made fresh daily.
Background	Akt, also referred to as PKB or Rac, plays a critical role in controlling cell survival and apoptosis (1-3). This protein kinase is activated by insulin and various growth and survival factors to function in a wortmannin-sensitive pathway involving PI3 kinase (2,3). Akt is activated by phospholipid binding and activation loop phosphorylation at Thr308 by PDK1 (4) and by phosphorylation within the carboxy terminus at Ser473. The previously elusive PDK2 responsible for phosphorylation of Akt at Ser473 has been identified as mammalian target of rapamycin (mTOR) in a rapamycin-insensitive complex with rictor and Sin1 (5,6). Akt promotes cell survival by inhibiting apoptosis through phosphorylation and inactivation of several targets, including Bad (7), forkhead transcription factors (8), c-Raf (9), and caspase-9. PTEN phosphatase is a major negative regulator of the PI3K/Akt signaling pathway (10). LY294002 is a specific PI3 kinase inhibitor (11). Another essential Akt function is the regulation of glycogen synthesis through phosphorylation and inactivation of GSK-3 α and β (12,13). Akt may also play a role in insulin stimulation of glucose transport (12). In addition to its role in survival and glycogen synthesis, Akt is involved in cell cycle regulation by preventing GSK-3 β -mediated phosphorylation and degradation of cyclin D1 (14) and by negatively regulating the cyclin-dependent kinase inhibitors p27 Kip1 (15) and p21 Waf1/Cip1 (16). Akt also plays a critical role in cell growth by directly phosphorylating mTOR in a rapamycin-sensitive complex containing raptor (17). More importantly, Akt phosphorylates and inactivates tuberin (TSC2), an inhibitor of mTOR within the mTOR-raptor complex (18,19).
Background References	1. Franke, T.F. et al. (1997) <i>Cell</i> 88, 435-7. 2. Burgering, B.M. and Coffer, P.J. (1995) <i>Nature</i> 376, 599-602. 3. Franke, T.F. et al. (1995) <i>Cell</i> 81, 727-36. 4. Alessi, D.R. et al. (1996) <i>EMBO J</i> 15, 6541-51. 5. Sarbassov, D.D. et al. (2005) <i>Science</i> 307, 1098-101. 6. Jacinto, E. et al. (2006) <i>Cell</i> 127, 125-37. 7. Cardone, M.H. et al. (1998) <i>Science</i> 282, 1318-21.

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#7142 PathScan[®] Total Akt1 Sandwich ELISA Antibody Pair



ELISA Antibody Pair

A. Solutions and Reagents

NOTE: Prepare solutions with reverse osmosis deionized (RODI) or equivalent grade water.

- 1. 20X Phosphate Buffered Saline (PBS): (#9808) To prepare 1 L 1X PBS: add 50 ml 20X PBS to 950 ml dH₂O,
- mix. 2. Wash Buffer: 1X PBS/0.05% Tween[®] 20, (20X PBST #9809).
- 3. Blocking Buffer: 1X PBS/0.05% Tween[®] 20, 1% BSA.
- 4. **1X Cell Lysis Buffer**: 10X Cell Lysis Buffer (#9803): To prepare 10 ml of 1X Cell Lysis Buffer, add 1 ml of 10X Cell Lysis Buffer to 9 ml of dH₂O, mix. Buffer can be stored at 4°C for short-term use (1–2 weeks).

Recommended: Add 1 mM phenylmethylsulfonyl fluoride (PMSF) (#8553) immediately before use.

- 5. Bovine Serum Albumin (BSA): (#9998).
- 6. TMB Substrate: (#7004).
- 7. STOP Solution: (#7002)

NOTE: Reagents should be made fresh daily.

B. Preparing Cell Lysates

For adherent cells

- 1. Aspirate media when the culture reaches 80–90% confluence. Treat cells by adding fresh media containing regulator for desired time.
- 2. Remove media and rinse cells once with ice-cold 1X PBS.
- 3. Remove PBS and add 0.5 ml ice-cold 1X Cell Lysis Buffer plus 1 mM PMSF to each plate (10 cm diameter) and incubate the plate on ice for 5 min.
- 4. Scrape cells off the plate and transfer to an appropriate tube. Keep on ice.
- 5. Sonicate lysates on ice.
- 6. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

For suspension cells

- 1. Remove media by low speed centrifugation (\sim 1,200 rpm) when the culture reaches 0.5–1.0 x 10⁶ viable cells/ml. Treat cells by adding fresh media containing regulator for desired time.
- 2. Collect cells by low speed centrifugation (~1,200 rpm) and wash once with 5-10 ml ice-cold 1X PBS.
- 3. Cells harvested from 50 ml of growth media can be lysed in 2.0 ml of 1X cell lysis buffer plus 1 mM PMSF.
- 4. Sonicate lysates on ice.
- 5. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

C. Coating Procedure

- 1. Rinse microplate with 200 μ l of dH₂O, discard liquid. Blot on paper towel to make sure wells are dry.
- 2. Dilute capture antibody 1:100 in 1 \tilde{X} PBS. For a single 96 well plate, add 100 µl of capture antibody stock to 9.9 ml 1X PBS. Mix well and add 100 µl/well. Cover plate and incubate overnight at 4°C (17–20 hr).

3. After overnight coating, gently uncover plate and wash wells:

- 1. Discard plate contents into a receptacle.
- 2. Wash four times with wash buffer, 200 µl each time per well. For each wash, strike plates on fresh paper towels hard enough to remove the residual solution in each well, but do not allow wells to completely dry at any time.
- 3. Clean the underside of all wells with a lint-free tissue.
- 4. Block plates. Add 150 μl of blocking buffer/well, cover plate, and incubate at 37°C for 2 hr.
- 5. After blocking, wash plate (Section C, Step 3). Plate is ready to use.

D. Test Procedure

- 1. Lysates can be used undiluted or diluted in blocking buffer. 100 μ l of lysate is added per well. Cover plate and incubate at 37°C for 2 hr.
- 2. Wash plate (Section C, Step 3).
- 3. Dilute detection antibody 1:100 in blocking buffer. For a single 96 well plate, add 100 μ l of detection antibody Stock to 9.9 ml of blocking buffer. Mix well and add 100 μ l/well. Cover plate and incubate at 37°C for 1 hr.
- 4. Wash plate (Section C, Step 3).
- 5. Secondary antibody, either streptavidin anti-mouse or anti-rabbit-HRP, is diluted 1:1000 in blocking buffer. For a single 96 well plate, add 10 µl of secondary antibody stock to 9.99 ml of blocking buffer. Mix well and add 100 µl/well. Cover and incubate at 37°C for 30 min.
- 6. Wash plate (Section C, Step 3).
- 7. Add 100 μl of TMB substrate per well. Cover and incubate at 37°C for 10 min.
- 8. Add 100 µl of STOP solution per well. Shake gently for a few seconds.
- 9. Read plate on a microplate reader at absorbance 450 nm.
 - 1. Visual Determination: Read within 30 min after adding STOP solution.
 - 2. **Spectrophotometric Determination**: Wipe underside of wells with a lint-free tissue. Read absorbance at 450 nm within 30 min after adding STOP solution.

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