

Primer Anneal/Extension

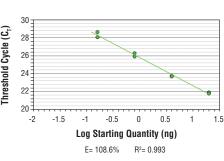
65°C



**Species Cross-Reactivity** 

Applications

ChIP	н	
Description: SimpleChIP® H contain a mix of forward and re specific to the intron 2 region of gene. These primers can be us been isolated using chromatin Primers have been optimized for titative real-time PCR and have with SimpleChIP® Enzymatic C #9003 and ChIP-validated anti Technology®. Dystrophin is a c that is responsible for the com muscle fibers to the extracellul sion of dystrophin is one of the dystrophy.	everse PCR primers that are of the human dystrophin sed to amplify DNA that has immunoprecipitation (ChIP). or use in SYBR® Green quan- e been tested in conjunction Chromatin IP Kits #9002 and ibodies from Cell Signaling critical protein in a complex nection of the cytoskeleton of lar matrix. Aberrant expres-	Sii Dhreshold Cycle (C.)



PCR Product Length

66 bp

impleChIP® Human DMD Intron 2 Primers were tested on NA isolated from cross-linked cells using the SimpleChIP $^{\scriptscriptstyle \otimes}$ Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. Real-time PCR was performed in duplicate on a serial dilution of 2% total input DNA (20 ng, 4 ng, 0.8 ng, and 0.16 ng) using a real-time PCR detection system and SYBR® Green reaction mix. The PCR amplification efficiency (E) and correlation coefficient (R<sup>2</sup>) were calculated based on the corresponding threshold cycle ( $C_{\tau}$ ) of each dilution sample during 40 cycles of real-time PCR (95°C denaturation for 15 sec, 65°C anneal/extension for 60 sec).

Storage: Supplied in nuclease-free water at a concentration of  $5 \,\mu$ M (each primer is at a final concentration of  $5 \,\mu$ M). Store at -20°C.

## Directions for Use:

1. Label the appropriate number of PCR tubes or PCR plates compatible with the model of real-time PCR machine to be used. PCR reactions should be performed in duplicate and should include a tube with no DNA to control for contamination, and a serial dilution of a 2% total input chromatin DNA (undiluted. 1:5, 1:25, 1:125), which is used to create a standard curve and determine amplification efficiency.

2. Add 2  $\mu$ l of the appropriate ChIP DNA sample to each tube or well of the PCR plate.

3. Prepare a master PCR reaction mix as described below. Add enough reagents for two extra reactions to account for loss of volume. Add 18  $\mu$ l of the master PCR reaction mix to each PCR reaction tube or well of the PCR plate.

Reagent	Volume for 1 PCR Reaction (20 µl)
Nuclease-free H <sub>2</sub> O	6 µl
5 µM SimpleChIP <sup>®</sup> Primers	
2X SYBR® Green React	ion Mix 10 µl

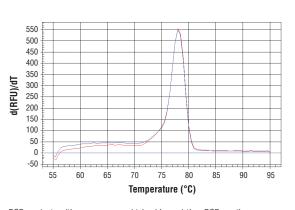
4. Start the following PCR reaction program:

- a. Initial Denaturation: 95°C for 3 min.
- b. Denaturation: 95°C for 15 sec.
- c. Anneal and Extension: Primer-specific temp. for 60 sec.
- d. Repeat steps b and c for a total of 40 cycles.

5. Analyze quantitative PCR results using software provided with the real-time PCR machine.



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PCR product melting curves were obtained for real-time PCR reactions performed using SimpleChIP® Human DMD Intron 2 Primers. Data is shown for both duplicate PCR reactions using 20 ng of total DNA. The melt curve consists of 80 melt cycles, starting at 55°C with increments of 0.5°C per cycle. Each peak is formed from the degradation of a single PCR product.

Applications Kev: W—Western IP—Immunoprecipitation IHC—Immunohistochemistry ChIP—Chromatin Immunoprecipitation IF-Immunofluorescence F-Flow cytometry E-P-ELISA-Peptide Species Cross-Reactivity Kev: H—human M—mouse R—rat Hm—hamster Mk—monkev Mi—mink C—chicken Dm—D. melanogaster X—Xenopus Z—zebrafish B-bovine All-all species expected Species enclosed in parentheses are predicted to react based on 100% homology.

Dg-dog Pg-pig Sc-S. cerevisiae Ce-C. elegans Hr-horse